


Designation: Meth A sarcoma

 CLS order number: Cryovial: 400284
 Vital: 440284

Origin and General Characteristics	
Depositor:	CLS
Organism:	Mouse (<i>Mus musculus</i>)
Strain:	Balb/c
Age:	Adult
Gender:	Female
Tissue:	Fibrosarcoma
Morphology:	Round cells
Cell type:	Infinitely growing tumor cell line
Growth Properties:	Suspension
Description:	Meth A sarcoma cells have been established as <i>in vitro</i> cell line from the transplantable Meth A sarcoma tumor, which was induced in 1962 using 3-methylcholanthrene by Old et al. in Balb/c mice.
References:	Old, Lloyd J., Boyse, Edward A., Clarke, Donald A., Carswell, Elizabeth A. Antigenic properties of chemically induced tumors. <i>Annals of the New York Academy of Sciences</i> 101(1): 80-106, 1962.
Culture Conditions and Handling	
Culture Medium:	RPMI 1640 medium supplemented with 2mM glutamine and 10% fetal bovine serum (MG-70, CLS order number 820700).
Subculturing:	Allow cell aggregates to settle to the bottom of the flask, discard the supernatant medium, disperse the cells with gentle pipetting and dispense into new flasks. Resuspend cell suspension in the flask and take representative aliquot to count the cell number per ml. Dilute cell suspension to 1×10^5 cells/ml with fresh medium and transfer into new flasks.
Split Ratio:	A ratio of 1:4 to 1:8 is recommended
Seeding density:	Start new cultures using $2-3 \times 10^5$ cells/ml; once the cells have recovered from the freezing and thawing process after 1-2 passages, adjust the cell density to 1×10^5 cells/ml when splitting the cells.
Fluid Renewal:	Every 2 to 4 days
Doubling time:	About 28 to 30 hrs
Freeze Medium:	CM-1 (CLS order number: 800125, 25ml, 800150, 50ml)
Freezing recovery:	About 53% of the initial cell number was collected after freezing.
Sterility:	Fluorescence (DAPI) test: negative; Mycoplasma specific PCR: negative; Bacteria specific PCR: negative
Biosafety Level:	1
Safety precautions:	If the cryovial is planned to be stored in liquid nitrogen and to be thawed in the future, special safety precautions should be followed: Protective gloves and clothing should be used and a facemask or safety goggles must be worn when transferring frozen samples into or removing from the liquid nitrogen tank. The removal of a cryovial from liquid nitrogen may result in the explosion of the frozen vial creating flying fragments. Caputo, J.L. Biosafety procedures in cell culture. <i>J. Tissue Cult. Methods</i> 11:223-227, 1988. ATCC

	Quality Control Methods for Cell Lines, 2nd edition, 1992.
Special Features of the Cell Line	
Tumorigenic:	Yes
Viruses:	SMRV: Negative, as confirmed by Real-Time PCR

Certificate of Analysis:	The Certificate of Analysis for each batch can be requested by e-mail at service@clsgmbh.de .
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Recommendations for handling of cells growing in suspension following delivery	
Cryopreserved cells	<p>If immediate culturing is not intended, the cryovial(s) must be stored below -150°C or at least at -80°C after arrival.</p> <p>If immediate culturing is intended, please follow these instructions:</p> <p>Quickly thaw by rapid agitation in a 37°C water bath within 40-60 seconds. The water bath should have clean water containing an antimicrobial agent. As soon as the sample has thawed, remove the cryovial from the water bath. Note: A small ice clump should still remain and the vial should still be cold.</p> <p>From now on, all operations should be carried out under aseptic conditions.</p> <p>Transfer the cryovial to a sterile flow cabinet and wipe with 70% alcohol. Carefully open the vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of culture medium (room temperature). Resuspend the cells carefully. Centrifuge at 300xg for 3 min and discard the supernatant. The centrifugation step may be omitted, but in this case the remains of the freeze medium have to be removed 24 hours later.</p> <p>Resuspend the cells carefully in 10ml fresh cell culture medium and transfer them into one T25 cell culture flask. All further steps are described in the Subculture section.</p>
Proliferating Cultures	<p>The cell culture flask, 1xT25, comes filled with cell culture medium.</p> <p>Incubate at 37°C for a minimum of 24 hrs.</p> <p>Count the cells, spin down the cell suspension at 300x g for 3 minutes to collect the cells. Resuspend the cells in an appropriate amount of fresh cell culture medium and transfer to new cell culture flasks.</p> <p>Incubate at 37°C for a minimum of 24 hrs.</p>

Warranty:	CLS warrants for a high cell viability and culture performance only if the product(s) is (are) stored and cultured according to the information described above. Using cell culture media and supplements other than the ones recommended in this product information may result in satisfactory proliferation and viabilities. CLS, however, does not warrant for cell recovery, proliferation and function if differing formulations are employed.
Disclaimer:	The customer shall not be entitled to employ this product for purposes other than research. Commercial utilization shall not be permitted; in particular, the cell line, its components or materials made therefrom shall not be sold or transferred to any third party. In addition, the term 'Commercial use' shall mean any activity by a party for consideration and may include, but is not limited to, use of the product or its components in manufacturing, for providing services, e.g. fee for service testing, in quality control or assurance processes within the manufacturing of products for sale, for therapeutic, diagnostic or prophylactic purposes, or for resale.